

Research article

Open Access

Identifying candidate genes affecting developmental time in *Drosophila melanogaster*: pervasive pleiotropy and gene-by-environment interaction

Julián Mensch*, Nicolás Lavagnino, Valeria Paula Carreira, Ana Massaldi, Esteban Hasson and Juan José Fanara

Address: Departamento de Ecología, Genética y Evolución, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Argentina

Email: Julián Mensch* - jmensch@ege.fcen.uba.ar; Nicolás Lavagnino - nlavagnino@ege.fcen.uba.ar;

Valeria Paula Carreira - vpcarreira@ege.fcen.uba.ar; Ana Massaldi - anamassaldi@hotmail.com; Esteban Hasson - ehasson@ege.fcen.uba.ar;

Juan José Fanara - jjfanara@ege.fcen.uba.ar

* Corresponding author

Published: 8 August 2008

Received: 29 February 2008

BMC Developmental Biology 2008, **8**:78 doi:10.1186/1471-213X-8-78

Accepted: 8 August 2008

This article is available from: <http://www.biomedcentral.com/1471-213X/8/78>

© 2008 Mensch et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background: Understanding the genetic architecture of ecologically relevant adaptive traits requires the contribution of developmental and evolutionary biology. The time to reach the age of reproduction is a complex life history trait commonly known as developmental time. In particular, in holometabolous insects that occupy ephemeral habitats, like fruit flies, the impact of developmental time on fitness is further exaggerated. The present work is one of the first systematic studies of the genetic basis of developmental time, in which we also evaluate the impact of environmental variation on the expression of the trait.

Results: We analyzed 179 co-isogenic single *P[GT1]*-element insertion lines of *Drosophila melanogaster* to identify novel genes affecting developmental time in flies reared at 25°C. Sixty percent of the lines showed a heterochronic phenotype, suggesting that a large number of genes affect this trait. Mutant lines for the genes *Merlin* and *Karl* showed the most extreme phenotypes exhibiting a developmental time reduction and increase, respectively, of over 2 days and 4 days relative to the control (a co-isogenic *P*-element insertion free line). In addition, a subset of 42 lines selected at random from the initial set of 179 lines was screened at 17°C. Interestingly, the gene-by-environment interaction accounted for 52% of total phenotypic variance. Plastic reaction norms were found for a large number of developmental time candidate genes.

Conclusion: We identified components of several integrated time-dependent pathways affecting egg-to-adult developmental time in *Drosophila*. At the same time, we also show that many heterochronic phenotypes may arise from changes in genes involved in several developmental mechanisms that do not explicitly control the timing of specific events. We also demonstrate that many developmental time genes have pleiotropic effects on several adult traits and that the action of most of them is sensitive to temperature during development. Taken together, our results stress the need to take into account the effect of environmental variation and the dynamics of gene interactions on the genetic architecture of this complex life-history trait.

Background

Development is a variable and time-dependent process that can be thought of as mapping genotypes on to adult phenotypes [1,2]. The time elapsed from the embryo to the reproductive phase, commonly known as developmental time (DT), is a trait of great relevance to fitness in all organisms [3]. In particular, *Drosophila* species occupy ephemeral habitats, such as rotting fruits that may result in selection for rapid development. Quoting Gould's *Ontogeny and Phylogeny*: "The timing of maturation is a primary variable in setting life history strategies. We have a prima facie case for ascribing direct significance to the change in developmental timing itself, not only to its morphological consequences" [4]. In addition, changes in the timing of developmental processes – heterochrony – may partly account for many evolutionary changes we observe because of genetic correlations of DT with other life-history related traits. Incidentally, heterochrony can also proceed by truncation in addition to retardation/acceleration of developmental processes [5]. The genes that control development are broadly shared among distantly related groups, and some of the pathways are surprisingly similar in superficially different organisms [6,7]. In this sense, experimental studies of egg-to-adult heterochrony at the molecular level can provide insights into the evolutionary processes by uncovering the genetic basis of ecologically relevant adaptive traits.

DT is a complex trait that displays considerable genetic variation in natural populations [8,9]. Laboratory selection for reduced DT triggered a negatively correlated response of other life-history traits, such as adult weight at eclosion [10], adult size [10-13], pre-adult survival [12,13] and longevity [14]. The reduction of adult fitness as a consequence of the acceleration of DT is a pattern called *fast development syndrome* and it illuminates the direct connection of the pre-adult and adult stages through energetic trade-offs [14]. These studies highlight the negative genetic correlations between DT and a large number of adult life-history traits, suggesting that genes involved in early development may also have pleiotropic effects on adult traits. However, specific genes contributing to variation in DT, and its correlations with other life-history traits, remain largely unknown.

The analysis of the genetic architecture of a trait requires not only the identification of the set of genes involved in its expression, but also their response to environmental variation (phenotypic plasticity) [15-17]. Phenotypic plasticity refers to the ability of a genotype to produce alternative morphological, behavioral and physiological characteristics in response to environmental conditions [18-21]. Temperature is an important environmental factor that affects the developing organism and surely has played a major role during the evolution of developmen-

tal traits in ectotherms [22-24]. Indeed, ectotherms must adapt their developmental program to a wide range of environmental temperatures during the day and also in different seasons along the year.

Drosophila brings an impressive toolkit for dissecting multiple interacting loci with individually small and environmentally sensitive effects that affect complex traits [15]. *P*-element mutagenesis is an effective strategy for studying the effects of single mutations on complex phenotypes [15]. A comprehensive understanding of DT requires the knowledge of: (1) the identities of the genes involved in the expression of the phenotype, (2) the underlying biological processes and molecular pathways, (3) the pleiotropic effects on adult traits and (4) the genetic basis of phenotypic plasticity in response to environmental variation.

Here we report a study of DT that reveals a large number of heterochronic mutants, most of which retard the time to reach the adult stage. The fastest developing line in our survey is a mutant of *Merlin*, a gene encoding a protein that regulates cell proliferation in developing imaginal discs. Interestingly, *Merlin* is the homolog of the human tumor suppressor gene *Neurofibromatosis Type 2* (NT2) involved in deregulation of cell proliferation in pathologies of the central nervous system [25]. In addition, we provide evidence for extensive pleiotropic effects of growth control genes on adult traits. Finally, we discovered several candidate genes involved in the plastic response to temperature variation.

Results

Pattern of variation among co-isogenic P[GT1] insertion lines

Substantial phenotypic variation among lines was observed for DT (Figure 1), wherein the most common effect was an increase of DT relative to the control. This is not unexpected as the most likely effect of mutations on a fitness-related trait is reasonable to be deleterious, although, we found a significant positive correlation ($r: 0.3, p < 3 \times 10^5$) between DT and viability [see Additional file 1]. This pattern revealed that the result observed of the heterochronic effect of the *P[GT1]* insertion lines can not be explained as a consequence of unfit flies. The most extreme phenotypes were exhibited by lines BG01543 and BG01412. The former developed 60 and 50 hours (males and females, respectively) faster and the latter 119 and 146 hours (males and females, respectively) slower than the control. We observed a significant line effect indicating mutational variance among single *P*-element insertion lines (Table 1). Indeed, the line effect accounts for 84% of total variance, five times higher than the error. In the ANOVA, differences between sexes were also significant, with males developing faster than females. In addition,

Table 1: Analysis of variance of mutational effects on developmental time.

| Source | d.f. | F | P | σ^2 (%) |
|-------------------|------|-------|---------|----------------|
| Line | 178 | 54.68 | <0.0001 | 84 |
| Sex | 1 | 46.87 | <0.0001 | Fixed |
| Line \times Sex | 178 | 0.76 | 0.99 | 0 |
| Error | 1066 | | | 16 |

d.f.: degree of freedom; σ^2 : component variance.

the contribution of the line-by-sex interaction to the total phenotypic variance was not significant.

Identification of co-isogenic P[GTI] insertion lines affecting DT and functional analysis

Dunnett analyses revealed that 107 out of 179 lines (60%) differed significantly from the control. Mean values of all significant P[GTI] insertion lines tested at 25°C are shown in Additional file 2. In seventy-four lines (40%) the insertion of the P-element caused an increase in DT, while in the rest of the significant lines DT was shortened as compared to the control (20%) (Figure 1). Those lines that exhibited significant DT differences relative to the control (heterochronic mutants) were considered as lines bearing an insertion in a candidate DT gene. Among fast developing lines, BG01543 and BG01902 stand out, since DT was reduced in each by more than 2 days. The P-element in line BG01902 disrupts *mastermind (mam)*, a gene encoding a glutamine-rich nuclear protein [26] that is a positive transcriptional regulator of the *Notch* signaling

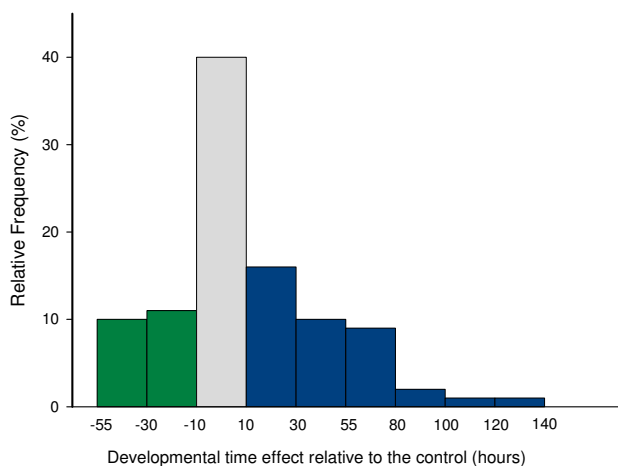


Figure 1
Frequency distribution of mutational effects on developmental time of 179 P[GTI] insertion lines.
 Grey bar represents lines showing similar effects relative to the control line (40%); green and blue bars represent lines showing a reduction (20%) and an increase (40%) in DT, respectively. Values in x-axis show the limits of each DT intervals.

pathway [27-30]. *Notch* is a central element in the cell signaling mechanism that controls a broad spectrum of cell fate choices during development across metazoans [31-33]. Interestingly, in humans, abnormalities in *Notch* signaling have been linked to a number of diseases, such as T cell acute lymphoblastic leukemia [34] and aortic valve disease [35]. In line BG01543, the P-element insertion occurs in *Merlin (Mer)*, a well-known negative regulator required for cell proliferation in developing imaginal discs of *Drosophila* [25]. It is a component of the *Hippo* signaling pathway, which is essential for the regulation of organ size during development [36]. *Mer* might control tissue growth by regulating endocytosis of membrane receptors in imaginal epithelia [37]. Interestingly, *Mer* is the homolog of the human tumor suppressor gene *Neurofibromatosis Type 2 (NF2)* involved in the de-regulation of cell proliferation in tumors of central nervous system pathologies [25,38].

Another gene involved in a cell proliferation pathway identified in our screen is *forkhead box, sub-group O (foxo)*. The insertion line BG01573 bearing a P-element in *foxo* increased DT by 24 hours in both sexes. *foxo* is a key regulatory component of the insulin-signaling pathway, which in *Drosophila* regulates the control of growth size of cells, organs, and the entire body in response to nutrient availability [39]. Puig *et al.* [40] established that *foxo* activates transcription in downstream as well as upstream targets of this signaling cascade by a transcriptional feedback mechanism that regulates cell growth and proliferation. It has been suggested that the insulin-signaling pathway regulates cell proliferation in imaginal discs, though the duration of the proliferation phases are controlled by Juvenile Hormone (JH) and ecdysteroids, that are themselves unaffected by the insulin-signaling pathway [41]. Premature ecdysone release leads to the rapid development of small adults, while the delay in ecdysone release extends developmental time and produces larger adults [42]. Recent studies have shown that the activity of the insulin-signaling pathway in the prothoracic gland modulates ecdysone release and influence both the length and the rate of larval growth [43-45]. It has been suggested that the neuropeptide *Amnesiac (Amn)* participates in regulating ecdysone synthesis in the prothoracic gland of *Drosophila* [43]. Interestingly, the P-element insertion mutant at *Amn* (line BG02286) also extended DT by more than two days in both sexes [see Additional file 2]. However, ecdysone can directly inhibit insulin and growth in the fat body and other peripheral tissues, an event that triggers metamorphosis. Furthermore, the ecdysone suppression of growth rate is lost in *foxo* mutants [45], indicating complex cross-talk between the ecdysone and the insulin signaling pathway during *Drosophila* development.

Our survey revealed extreme heterochronic mutants, particularly in the direction of increased DT [see Additional file 2]. For instance, the *P*-element insertion in the gene *Karl* (BG01010) extended DT by more than 100 hours in both sexes. The molecular function inferred from protein sequence suggests that *Karl* is a retinol-binding protein with no clear association to a biological process. In line BG00372 we registered a delay of 93 hours and 108 hours for males and females, respectively. In this case the *P*-element has inserted 1498 bp downstream of the 3' end of the *CG1678* gene. The molecular function and the biological processes in which this gene is involved are unknown. However, there is evidence that *CG1678* interacts with other genes such as *And*, *ewg*, *CG1472*, *rl*, *Bsg25D*, *CG11275* and *msb11* [46]. Line BG01412 exhibited the most extreme phenotype in our screen, the *P*-element insertion was associated with an extended developmental time of 119 and 146 hours in males and females, respectively. Unfortunately, information about the nucleotide sequences flanking the *P*-element insertion site is not available.

Candidate DT genes are involved in a wide range of biological processes according to their gene-ontology (GO) terms (Table 2). Interestingly, DT genes not only are involved in processes associated to organismal development but also to biosynthetic and cellular metabolic processes. We did not find significant differences between groups of candidate genes accelerating development vs those that extended development in the distribution among GO terms, suggesting that similar ontogenetic

Table 2: Distribution of candidate DT genes among biological process GO terms.

| Biological process GO terms | % of genes |
|--|------------|
| Cellular metabolic process | 57.9 |
| Multicellular organismal development | 39.5 |
| Macromolecule metabolic process | 36.8 |
| Anatomical structure development | 34.2 |
| Regulation of biological process | 31.6 |
| Cellular developmental process | 31.6 |
| Cell communication | 29.0 |
| Biosynthetic process | 23.7 |
| Cellular component organization and biogenesis | 18.4 |
| Sexual reproduction | 18.4 |
| Establishment of localization | 18.4 |
| Nitrogen compound metabolic process | 10.5 |
| Behavior | 10.5 |
| Response to stress | 10.5 |
| Cell proliferation | 10.5 |
| Cell adhesion | 10.5 |
| Cell cycle | 7.9 |

Genes are distributed in a non-exclusive manner (i.e. a given gene might be associated to more than one GO term, see text for more details). The percentage of genes related to each GO term is shown.

pathways may be responsible for both types of heterochronic phenotypes.

Candidate genes for plasticity

Growth and development of ectotherms are determined in part by their thermal environment [22,23]. In particular, temperature during ontogeny exerts a strong influence shaping the evolution of larval traits [24,47,48]. In fact, phenotypic responses result of adaptation to different thermal environments and/or may be an unavoidable consequence of the effect of temperature on the organism's physiology during development [49]. In this context, the pattern of phenotypic effects of *P*-element insertion lines reared at different developmental temperatures would provide new insights in the study of phenotypic evolution of larval traits. The ANOVA showed that differences among lines and between thermal treatments were significant (Table 3). More importantly, our screen revealed that the line-by-sex and the line-by-temperature interactions were also highly significant, indicating that the behavior of each line depended on the temperature at which it was reared and the sex. However, there was a large difference in the magnitude of these genotype-by-environment interactions. The former accounts for only 1% of the total phenotypic variance (a percentage similar to that obtained in the 25°C assay) while the line-by-temperature interaction explained 52% of the variation. Moreover, note that in the general assay, the percentage of total phenotypic variance explained by differences among lines was 84%, whereas in our assays of phenotypic plasticity this percentage dropped to 30%. It may be argued that part of the effect was obscured by the high value of the line-by-temperature interaction term. This observation opens an excellent opportunity for studying the genetic basis of phenotypic plasticity of developmental time. Thus, we decided to analyze whether the significant line-by-temperature interaction can be explained by changes in magnitude of among-line variance across thermal treatments or changes in the rank order among lines, i.e. a cross-temperature genetic correlation lower than

Table 3: Analysis of variance of mutational effects on developmental time among lines tested at different thermal treatments.

| Source | d.f. | F | P | σ^2 (%) |
|--------------------------------|------|------|---------|----------------|
| Line | 41 | 2.11 | 0.0085 | 30 |
| Sex | 1 | 8.82 | 0.0047 | Fixed |
| Thermal Treatment | 1 | 4.56 | 0.03 | Fixed |
| Line × Sex | 41 | 1.73 | 0.041 | 1 |
| Line × Thermal Treatment | 41 | 41.4 | <0.0001 | 52 |
| Sex × Thermal Treatment | 1 | 9.26 | 0.0038 | Fixed |
| Line × Sex × Thermal Treatment | 41 | 0.56 | 0.9886 | 0 |
| Error | 479 | | | 17 |

d.f.: degree of freedom; σ^2 component variance.

unity. Our results showed that about half of the interaction variance can be explained by a greater among-line variance observed at 17°C than 25°C and the other half by temperature-specific effects on DT of the lines (Figure 2).

Thirty out of 42 lines tested at 17°C and 25°C showed significant differences relative to the control at one or both temperatures (Table 4). In addition, DT measured at different temperatures were positive correlated for the set of 30 heterochronic lines in both sexes (males: $r: 0.61, p < 0.05$; females: $r: 0.55, p < 0.05$; Figure 3). These results imply that most of the mutations that affected DT at both temperatures did so in the same phenotypic direction (either increasing or decreasing DT). Forty seven percent of the lines showed this pattern (bottom left and top right quadrants in Figure 3). Nevertheless, we also identified a set of lines in which the effect of the mutation on DT was temperature-dependent, suggesting that the mutated loci may be possible candidates for temperature plasticity genes. Indeed, forty percent of the lines affected DT in only one of the temperatures. We observed three lines showing a heterochronic phenotype only at 25°C. The line bearing an insertion in *CG14478* is an example of this

pattern. In contrast, nine lines showed a heterochronic phenotype only at 17°C. For example, the insertion in *Imp* exhibited significant differences at 17°C in both sexes, but not at 25°C. Surprisingly, four lines (13%) affected DT in opposite phenotypic directions in the two rearing temperatures. On the one hand, line BG02159, bearing a mutation in *CG32666*, decreased and increased DT at 17°C and 25°C, respectively. On the other hand, mutant lines for genes *Nmdar*, *sugarless* and *CG11226* exhibited the opposite pattern (Figure 4).

Discussion

Genetic architecture of developmental time

The genetic architecture of a trait determines the variational properties of the phenotype, that is, its evolutionary potential. Our study revealed 116 heterochronic independent mutants involved in the expression of DT (107 lines showed a mutant phenotype at 25°C, while 9 lines were heterochronic only at 17°C). In this sense, we were capable to identify components of different kind of pathways involved in DT expression. On the one hand, some components correspond to molecular mechanisms directly involved in timekeeping processes, such as, the ecdysone and the insulin-signaling pathway [41,50,51]. It

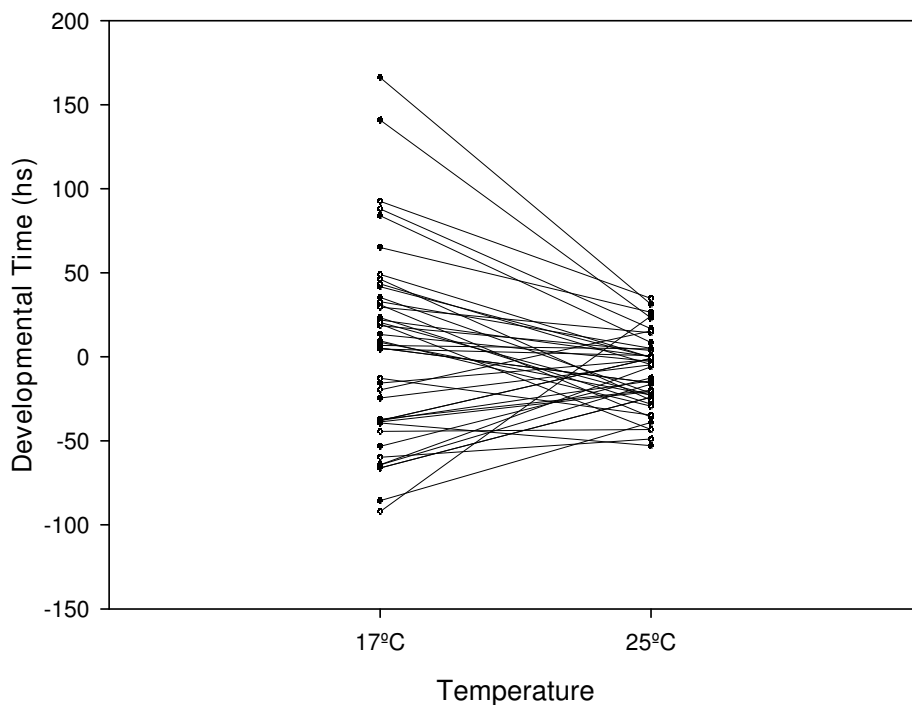


Figure 2
Reaction norms of 45 P[GTI] insertion lines tested at 17°C and 25°C. Values are shown as the deviation of the insert line mean from contemporaneous control line.

Table 4: Effects of significant *P[GT1]* insertion lines screened at different thermal treatments.

| Line | Gene | Males 17°C | Females 17°C | Males 25°C | Females 25°C | S | T | S × T |
|---------|----------------|------------|--------------|------------|--------------|-----|-----|-------|
| BG00369 | <i>CG13334</i> | -34.25* | -44.73** | -44.50*** | -56.99** | N | *** | N |
| BG00373 | <i>CG11226</i> | 84.18*** | 72.11*** | 16.22* | 23.52*** | N | *** | N |
| BG00386 | <i>Nmdar1</i> | NS | 54.81** | -30.73*** | -36.13*** | N | *** | * |
| BG00737 | <i>Hsp27</i> | -64.24*** | -55.94*** | NS | NS | ** | *** | N |
| BG00930 | no sequence | 45.08*** | NS | NS | NS | ** | *** | N |
| BG01028 | <i>CG33260</i> | -35.5* | -41.24** | NS | -21.81** | N | ** | N |
| BG01214 | <i>CG10064</i> | 47.86*** | NS | -16.82* | -28.59*** | *** | *** | N |
| BG01339 | <i>ctt</i> | 92.63*** | 93.64*** | 39.14*** | 29.46*** | N | *** | N |
| BG01354 | <i>CG30492</i> | NS | 41.53** | NS | NS | ** | *** | N |
| BG01488 | <i>msi</i> | NS | -41.18** | -19.40** | -26.76*** | N | * | N |
| BG01672 | <i>CG14591</i> | -85.53*** | -48.45** | -27.40** | NS | N | *** | N |
| BG01716 | <i>paps</i> | -38.8* | -39.38* | NS | -22.74** | * | *** | N |
| BG01726 | <i>CG11382</i> | -39.82** | -38.27* | NS | NS | N | *** | N |
| BG01735 | <i>bib</i> | NS | NS | NS | -19.91** | N | N | * |
| BG01763 | <i>CG33960</i> | 173*** | 166.28*** | 34.41*** | NS | N | ** | N |
| BG01780 | <i>CG11226</i> | 42.8** | 49.41*** | -20.18* | -32.31*** | N | *** | N |
| BG01822 | <i>Imp</i> | -47.95** | -52.38** | NS | NS | N | *** | N |
| BG01902 | <i>mam</i> | -59.92*** | -62.62*** | -47.64*** | -53.05*** | N | * | N |
| BG01912 | <i>pxb</i> | -79.85*** | -97.30*** | -33.10*** | -45.92*** | N | *** | N |
| BG01990 | <i>CG30492</i> | -43.82** | -43.86** | -35.87*** | -46.05*** | N | N | N |
| BG02042 | <i>eas</i> | 47.78** | NS | NS | NS | N | *** | N |
| BG02088 | <i>CG15309</i> | 49.55** | 46.28** | NS | NS | N | *** | N |
| BG02102 | <i>CG13434</i> | -61.79*** | -70.34*** | -16.73* | -17.89* | * | *** | N |
| BG02157 | <i>CG8177</i> | 90.67*** | 76.60*** | NS | NS | N | *** | N |
| BG02159 | <i>CG32666</i> | -85.12*** | -96.73*** | 29.51*** | NS | N | *** | * |
| BG02239 | <i>CG11550</i> | 63.36*** | 60.57** | 29.40*** | 22.72*** | N | *** | N |
| BG02462 | <i>CG6301</i> | 50.77*** | 38.37* | NS | NS | * | *** | N |
| BG02690 | <i>CG14478</i> | NS | NS | -32.02*** | -37.68*** | N | *** | N |
| BG02747 | <i>rut</i> | 136.73*** | 129.40*** | 28.00*** | NS | N | *** | N |
| BG02823 | <i>scyl</i> | NS | NS | 17.44* | NS | N | *** | N |

Values are shown as the deviation of the insert line mean from contemporaneous control line. Asterisks in columns S, T and S × T represent the quantitative effect of Sex, Thermal treatment and Sex-by-Thermal treatment in response to environmental variation (see materials and methods for detail statistical treatment). **p* < 0.05; ***p* < 0.01; ****p* < 0.001; NS: no significant.

has been established that environmental inputs during ontogeny influences multiple steps of these kind of heterochronic gene pathways, such as, the nutrition status and the light: dark regimes. A paradigmatic example of this kind of environmental input is the circadian rhythm *period* mutants that alter DT in *Drosophila* [52]. On the other hand, we also observed heterochronic phenotypes that seem to be a by-product of the disruption of a pathway that is not explicitly involved in the control of temporal process but plays a major role in organ growth. This is the case of *Merlin*, a negative regulator of the *Hippo* signaling pathway, required for cell proliferation in developing imaginal discs. Interestingly, *Merlin* is a component of a tumour-suppressor network associated with human tumorigenesis [36]. All in all, our results support the hypothesis raised by Moss [53] that timing of development is control directly and indirectly by different ontogenetic pathways.

Since the same co-isogenic lines were also screened for abdominal and sternopleural bristle number, starvation

resistance and olfactory behavior [17,54,55] we also examined whether DT candidate genes have pleiotropic effects on adult fitness-related traits. Several DT heterochronic lines (genes) affect starvation resistance (4 genes), bristle number (8 genes) and olfactory behavior (13 genes) [see Additional file 3]. Once again, *Merlin* seems to play a role not only in egg-to-adult DT but also in starvation resistance and adult olfactory behavior. In BG01543 (*Mer* mutant), the *P[GT1]* insertion in exon 1 caused a reduction in the RNA expression levels in embryos, third instar larvae and adults but not in the pupal stage compared to the control [17] indicating that anomalies caused by the disruption of *Merlin* on DT took place before metamorphosis. Since insects do not grow as adults, their final size is a product of growth rate and length at larval stages [56]. Surprisingly, the mutated *Merlin* line did not differ significantly from the control in several morphological traits such as face width, head width, thorax length and wing size suggesting a decoupled mechanism of timing control and size development (Carreira, Mensch & Fanara, unpublished results). Regarding the pleiotropic

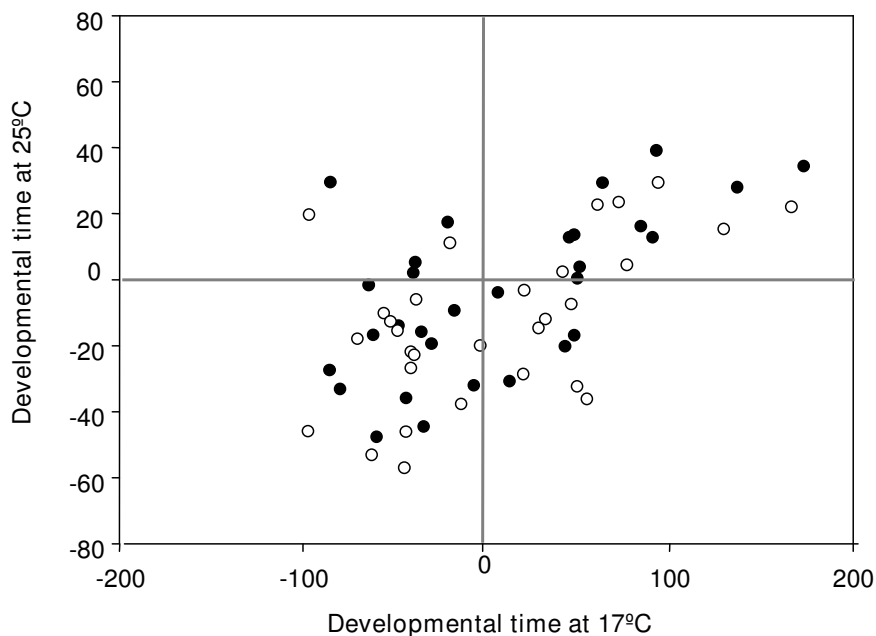


Figure 3
Correlation of 30 *P[GTI]* heterochronic mutants tested at 17°C and 25°C. Values are shown as the deviation of the insert line mean from contemporaneous control line for each sex separately; males (solid circles) and females (open circles).

effect of *Mer* on starvation resistance and adult olfactory behavior, it is not easy to discern whether aberrant phenotypes are the consequence of early developmental events or are directly related to the functional disruption of the gene product in the adult. As another example, the insertion line for *tramtrack* (*ttk*), a transcription repressor gene involved in cell-fate determination [57], increased DT by two days in both sexes and also affected starvation resistance and olfactory behavior. These interesting examples constitute clear evidence of an intricate network of pleiotropic effects of key developmental genes throughout the life-cycle of a holometabolous organism. Such pervasive pleiotropy of key genes that control development, affecting early and late fitness-related traits, show an integrated picture of the evolution of life-history traits at the molecular level.

Regarding the effect of sex, males developed significantly faster than females, an observation that is quite striking since it is at odds with extensive literature showing that *Drosophila* females usually reach adulthood before males in the species studied so far [58]. Moreover, a recent study by Paranjpe *et al.* [59] showed that females developed faster than males in *D. melanogaster*. However, some differences between our study and Paranjpe *et al.*'s in terms of the experimental design are worth mentioning. Paranjpe *et al.* found that females developed faster than males under several photoperiod regimes during ontogeny,

including 12:12 L:D (the regime performed in our study). Nevertheless, we must note that different genetic backgrounds were analyzed in these two studies. Indeed, our control line, which shares the same genetic background with all mutant lines tested, did not show any sign of sexual dimorphism in DT, stressing the need to take into account the effect of the genetic background on the expression of the analyzed trait since genetic background \times gene interactions are known to be quite pervasive [60,61]. At the same time the null input of the line-by-sex interaction to the total phenotypic variance adds a complementary aspect to the discussion of the absence of sexual dimorphism for DT. In the last few years a similar set of coisogenic insertion lines was studied for several adult traits for which the line-by-sex interaction explained significant portions of variance. In this context, our DT survey yielded the lowest value in this category, followed by sternopleural bristle number (5%), adult odor-behavior (13%), abdominal bristle number (21%) and starvation resistance (66%) [17,54,55]. Among the heterochronic mutants, nine had a male-specific effect and in seven only female DT was affected. It is important to note that none of the lines showed opposite phenotypic effects across sexes, meaning that we did not find any lines that decreased DT in one sex and increased the trait in the other. In conclusion, our results indicate that there are features of the genetic architecture of DT that are highly conserved in both sexes.

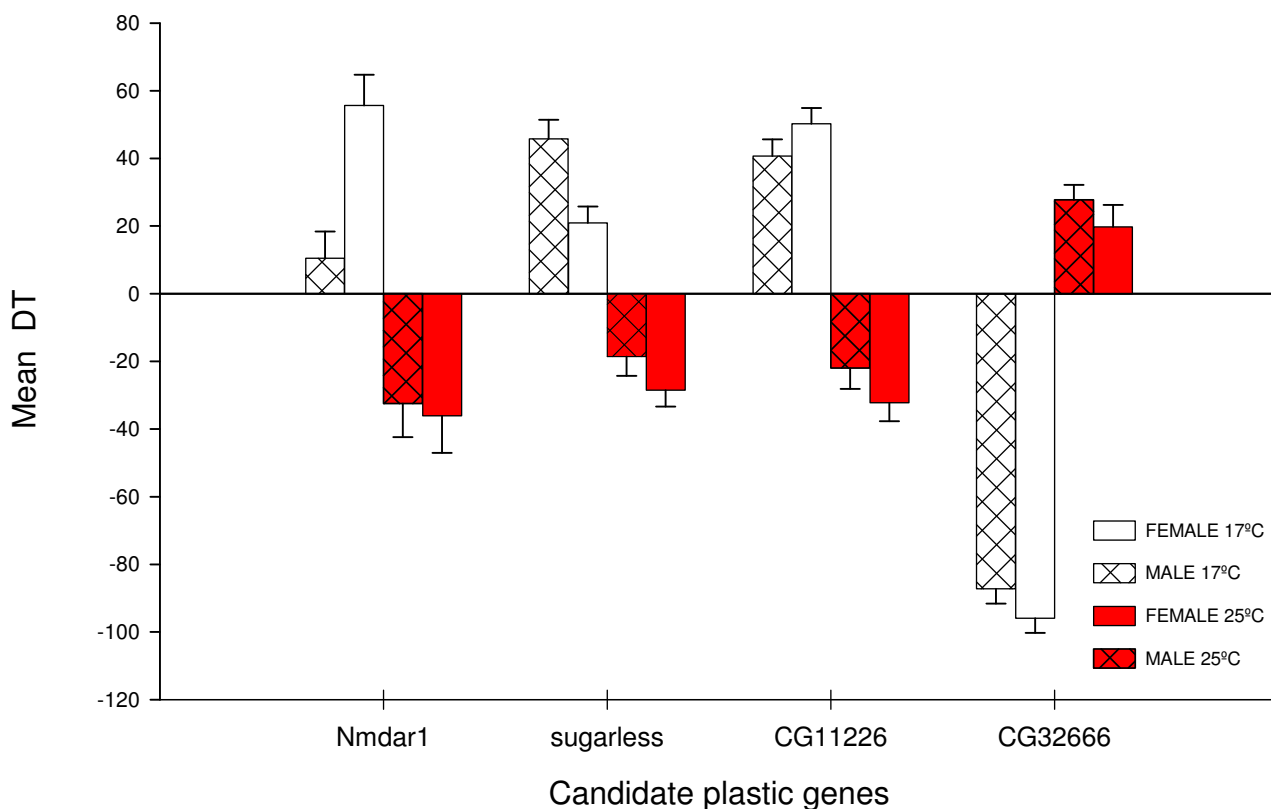


Figure 4
Heterochronic mutants having opposite phenotypic effects between thermal treatments. 17°C assay is represented with open bars and 25°C assay with red bars. Males are represented with striped bars and females with open bars.

Gene-by-environment interaction

Although the multiple interacting genes affecting complex traits can readily be dissected, how much genotype-environment interactions contribute to variation in these traits remains elusive. The quantitative dissection of developmental time reported here reveals that almost all heterochronic lines tested at different developmental temperature presented an effect with quantitative variation between temperatures, a remarkable consequence of the large gene-by-environment interaction (Table 4). The most striking examples of these developmental reaction norms are the mutant lines for genes *CG11226*, *CG32666*, *Nmdar1*, and *sugarless* (Figure 4). *CG32666* gene product is a receptor signaling protein with serine/threonine kinase activity, *NMDAR1* is a ionotropic glutamate receptor and *SUGARLESS* is a protein with UDP-glucose 6-dehydrogenase activity indicating that this routine cellular metabolic processes are not only involved in developmental time but also that are sensitive to environmental variation.

The part of the genetic diversity that has the potential to affect the phenotype, but that is not expressed under the current genotypic or environmental conditions, is referred to as "cryptic" or "hidden" genetic variation [62]. Molecular mechanisms responsible for this particular genetic variation include epistasis and genotype-by-environment interactions. In our case, in addition to the plastic developmental reaction norms, large input to the gene-by-environment interaction refers to the bigger variance magnitude at 17°C (Figure 2). It is possible that at 25°C genetic variation was buffered in comparison to the variation expressed at 17°C, an unusual rearing temperature for these lines. Taken together, our results highlight the large potential of the *Drosophila* genome to changes in this relevant environmental factor, although this genetic plasticity was exhibited by these particular inbred lines, a pattern that it is not necessarily true for outbred populations. Future efforts will focus on elucidating the molecular mechanisms controlling the temperature plastic response observed. Since in nature insects are exposed to a wide range of environmental temperatures during the day and also in different seasons along the year with local-specific

thermal regimes, we are also interested in studying the evolutionary forces shaping the variation in the candidate DT genes.

Conclusion

The study of heterochrony has played an important role in the intersection of evolution, genetics and developmental biology since the late nineteenth century. During the last decade the concept has been revitalized with studies at cellular and molecular level. Here we identified components of several integrated time-dependent pathways affecting egg-to-adult DT and also components of pathways that are not explicitly involved in the control of temporal processes. In this sense, we identified 116 egg-to-adult heterochronic mutants, most of them developed slower compared to the control. However, we identified a set of lines that developed faster by more than two days. This is the case of *Mer* mutant line, a gene involved in a cell proliferation pathway. *Mer* and others DT candidate genes have pleiotropic effects on adult traits as well. Most of developmental time candidate genes were sensitive to the rearing temperature, a fact that stresses the need to take into account the effect of environmental variation on the genetic architecture of complex traits.

Methods

Drosophila stocks and Developmental time assays

We scored 179 homozygous viable *P[GT1]* insertion lines, constructed in a co-isogenic *Canton-S B* background [63] as part of the *Berkeley Drosophila Genome Project* (see Availability and requirements section for URL) for DT.

Batches of 30–40 P-element insertion lines were assessed simultaneously. To account for environmental variation in DT between batches, 8 replicated vials of the control strain (a co-isogenic P-element insertion free line with the same genetic background) was run in parallel with each batch. For each line, 300 pairs of sexually mature flies were placed in oviposition chambers for 8 hours. Eggs were allowed to hatch and batches of 30 first-instar larvae were transferred to culture vials containing a cornmeal-agar-molasses medium (4 replicates vial per line). Emerged flies from each vial were collected every 12 hours and sorted by sex. We estimated DT as the time elapsed since the transfer of first-instar larvae to the vials until adult emergence. Each vial was kept in an incubator at $25^{\circ}\text{C} \pm 0.5$, under a 12:12 h light:dark photoperiod and at 60–70% of humidity. Thirteen lines with low viability (those that reached less than 50% of control pre-adult survival) were excluded from our survey to avoid the confusing effect of high pre-adult mortality with DT. A subset of 42 lines selected at random was also screened at $17 \pm 0.5^{\circ}\text{C}$.

Statistical analysis

Quantitative genetic analysis

Analysis of variance (ANOVA) was utilized to assess the magnitude of mutational variance for DT at each temperature. In order to include all lines tested in different batches, individual DT scores were expressed as deviations from the mean of their contemporaneous co-isogenic controls, separately for males and females. Three-way ANOVA was computed for each thermal treatment, following the mixed model: $Y = \mu + L + S + L \times S + E$, where μ is the overall mean, L is the random effect of line, S is the fixed effect of sex and E represents the error term. An ANOVA was used to assess the magnitude of mutational variance for DT induced by *P[GT1]* insertions. We also estimated the relative contribution of all random sources of variation (Line, Line \times Sex, and error) to the total variance.

To identify which lines were responsible of the significant line or line-by-sex interaction, phenotypic differences between *P*-element insertion lines and the control were tested using *Dunnnett* contrasts for each temperature and batch. Those lines that exhibited significant DT differences relative to the control (heterochronic mutants) were considered as lines bearing an insertion in a candidate DT gene.

Gene-by-thermal treatment analysis

In those lines screened at both temperatures (17 and 25°C) we studied the effect of phenotypic plasticity by thermal treatments following the mixed ANOVA model, $Y = \mu + L + S + T + L \times S + L \times T + S \times T + L \times S \times T + E$, where μ is the overall mean, L is the random effect of line, S is the fixed effect of sex, T is the fixed effect of thermal treatment and E represents the error term. Since significant $G \times E$ (genotype \times environment) can arise due to: (1) differences among- lines variance in separate environments (change in scale) and/or (2) deviations from unity of the cross-environment genetic correlation ($r_{G \times E} < 1$, see below) (changes in rank order), the contribution of these two sources of variation was analyzed by means of the equation derived by Robertson [64], $V_{G \times E} = [(\sigma_{E1} - \sigma_{E2})^2 + 2\sigma_{E1}\sigma_{E2}(1 - r_{G \times E})]/2$, where $V_{G \times E}$ is the $G \times E$ variance component, $r_{G \times E}$ is the cross-environment genetic correlation and, σ_{E1} and σ_{E2} are the square roots of the among-line variance components of the two thermal environments studied [65]. The first term corresponds to differences in among-line variance while the second corresponds to deviations from the perfect correlation across environments ($r_{G \times E} < 1$). The cross-environment genetic correlation ($r_{G \times E} < 1$) is the genetic correlation of measurements in different environments and here reflects the degree in which the same genes control the expression across environments. $r_{G \times E}$ was estimated as: $r_{G \times E} = \text{COV}_{E1E2}/\sigma_{E1}\sigma_{E2}$, where COV_{E1E2} is the covariance

of lines in different environments and it was calculated as the covariance of the lines in different environments.

Finally, we studied the phenotypic change by the effect of the *P* element insertion in each line showing significant differences with respect to the control in at least one thermal assay. In this case we used the following fixed ANOVA model:

$Y = \mu + T + S + T \times S + E$, where T stands for the effect of thermal treatment, S for the effect of sex and E represents the error term. Those lines that exhibited significant differences in DT between thermal treatments (T factor) were considered as lines bearing a mutation in a candidate gene involved in the plastic response of DT to temperature variation.

All statistical analyses were performed using STATISTICA software packages (StatSoft, Inc. 1999, 2001).

Gene identification and functional analysis

In order to identify the mutated genes, nucleotide sequences flanking the *P*-element insertion in each candidate line were used to search for homologous regions in the *D. melanogaster* genome. Searches of homologous sequences were performed against Release 5 of the published *D. melanogaster* genomic sequence using the Pubmed server. Flybase (see Availability and requirements section for URL) was used to identify target genes. Candidate DT genes were distributed in different biological processes according to their gene-ontology (GO) terms [66]. This analysis was performed automatically with the aid of the program FatiGO+, from the Babelomics suite of bioinformatic tools, available at (see Availability and requirements section for URL)[67]. This program distributes the genes among the GO terms in a non-exclusive manner (i.e. a given gene might be associated to more than one GO term). Using the same program we made a comparison of the distribution of GO terms between the sample of lines that developed faster and the population of lines that developed longer in order to find GO terms that were significantly over-represented in one of the samples.

Availability and requirements

Berkeley Drosophila Genome Project: <http://flypush.imgen.bcm.tmc.edu/pscreen/>

Flybase: <http://flybase.bio.indiana.edu/>

FatiGO+: <http://www.fatigo.org/>

Abbreviations

DT: developmental time; ANOVA: analysis of variance; GO: gene-ontology.

Authors' contributions

JM and JJF conceived the study. JM, NL, VPC and AM performed the developmental time assays. JM analyzed the data and wrote the first draft of this manuscript. JM, EH and JJF wrote the final version. NL and VPC helped to draft the final version of this manuscript. All authors read and approved the final manuscript.

Additional material

Additional file 1

Correlation between viability (VT) and developmental time (DT) for all P[GT1] insertion lines analyzed. VT is presented in angular transformation (arcsine of the square root of the proportion of survival) and DT expressed in hours. Solid line represents the correlation plot and dashed lines the 95% confidence intervals ($r = 0.305$).

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-213X-8-78-S1.bmp>]

Additional file 2

Mean values of significant P[GT1] insertion lines on DT (25°C). Values are shown as the deviation of the insertion line mean from contemporaneous control for each sex separately.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-213X-8-78-S2.doc>]

Additional file 3

Genetic information of candidate DT genes including gene names, P[GT1] insertion sites, cytological positions, biological process gene-ontologies and pleiotropy.

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1471-213X-8-78-S3.doc>]

Acknowledgements

The authors wish to thank T.F.C. Mackay and R. R. H. Anholt for comments and suggestions that helped to improve this article. We also want to thank A. Yamamoto for technical support, two anonymous reviewers for their constructive criticisms and all colleagues at the Laboratorio de Evolución. This work was supported by grants of the Universidad de Buenos Aires, Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET) and Agencia Nacional de Promoción Científica y Técnica. J. Mensch, N. Lavagnino and V.P. Carreira are recipients of post-graduate scholarships of CONICET. E.H. and J.J. Fanara are members of Carrera del Investigador Científico (CONICET).

References

1. Wallace A: **The emerging conceptual framework of evolutionary developmental biology.** *Nature* 2002, **415**:757-764.
2. Reed RD, Chen PH, Nijhout FH: **Cryptic variation in butterfly eyespot development: the importance of sample size in gene expression studies.** *Evol Dev* 2007, **9**:2-9.

3. Stearns SC: **The evolution of Life Histories.** Oxford University Press; 1992.
4. Gould SJ: **Ontogeny and phylogeny.** Harvard University Press; 1977.
5. Smith KK: **Time's arrow: heterochrony and the evolution of development.** *Int J Dev Biol* 2003, **47**:613-621.
6. Carrol S: *Endless forms most beautiful* W.W. Norton & Company; 2005.
7. Pires-daSilva A, Sommer RJ: **The evolution of signalling pathways in animal development.** *Nat Rev Genet* 2003, **4**:39-49.
8. Cortese MD, Norry FM, Piccinali R, Hasson E: **Direct and correlated responses to artificial selection on developmental time and wing length in *Drosophila buzzatii*.** *Evolution* 2002, **56**:2541-2547.
9. Fanara JJ, Folguera G, Iriarte PF, Mensch J, Hasson E: **Genotype by environment interactions in viability and developmental time in populations of cactophilic *Drosophila*.** *J Evol Biol* 2006, **19**:900-908.
10. Nunney L: **The response to selection for fast larval development in *Drosophila melanogaster* and its effect on adult weight: an example of a fitness trade-off.** *Evolution* 1996, **50**:1193-1204.
11. Zwaan BJ, Bijlsma R, Hoekstra RF: **Artificial selection for development time in *Drosophila melanogaster* in relation to the evolution of aging: direct and correlated responses.** *Evolution* 1995, **49**:635-648.
12. Chippindale AK, Alipaz JA, Chen H, Rose M: **Experimental evolution of accelerated development in *Drosophila*. 1. Developmental speed and larval survival.** *Evolution* 1997, **51**:1536-1551.
13. Prasad NG, Shakarad M, Anitha D, Rajamani M, Joshi A: **Correlated responses to selection for faster development and early reproduction in *Drosophila*: the evolution of larval traits.** *Evolution* 2001, **55**:1363-1372.
14. Chippindale AK, Alipaz JA, Rose MR: **Experimental evolution of accelerated development in *Drosophila*. 2. Adult fitness and the fast development syndrome.** In *Methuselah Flies: A case study in the evolution of aging* Edited by: Rose MR, Passananti HB, Matos M. Singapore: World Scientific; 2004:413-435.
15. Mackay TFC: **The genetic architecture of quantitative traits: lessons from *Drosophila*.** *Curr Opin Genet Dev* 2004, **14**:253-257.
16. Alonso JM, Ecker JR: **Moving forward in reverse: genetic technologies to enable genome-wide phenomic screens in *Arabidopsis*.** *Nat Rev Genet* 2006, **7**:524-536.
17. Sambandan D, Yamamoto A, Fanara JJ, Mackay TF, Anholt R: **Dynamic genetic interactions determine odor-guided behavior in *Drosophila melanogaster*.** *Genetics* 2006, **174**:1349-1363.
18. West-Eberhard MJ: **Developmental plasticity and evolution.** Oxford University Press; 2003.
19. Schlichting C, Pigliucci M: **Phenotypic Evolution: a reaction norm perspective.** Sinauer Associates; 1998.
20. Fordyce JA: **The evolutionary consequences of ecological interactions mediated through phenotypic plasticity.** *J Exp Biol* 2006, **209**:2377-2383.
21. Mackay TFC, Anholt R: **Ain't misbehavin'? Genotype-environment interactions and the genetics of behavior.** *Trends Genet* 2007, **23**:311-314.
22. Davidowitz G, Nijhout F: **The physiological basis of reaction norms: the interaction among growth rate, the duration of growth and body size.** *Integr Comp Biol* 2004, **44**:443-449.
23. Angilletta M, Steury T, Sears M: **Temperature, growth rate, and body size in ectotherms: fitting pieces of a life-history puzzle.** *Integr Comp Biol* 2004, **44**:498-509.
24. Trotta V, Calboli FC, Ziosi M, Guerra D, Pezzoli MC, David JR, Cavicchi S: **Thermal plasticity in *Drosophila melanogaster*: a comparison of geographic populations.** *BMC Evol Biol* 2006, **6**:67.
25. Hariharan IK, Bilder D: **Regulation of imaginal disc growth by tumor-suppressor genes in *Drosophila*.** *Annu Rev Genet* 2006, **40**:335-361.
26. Smoller D, Friedel C, Schmid A, Bettler D, Lam L, Yedvobnick B: **The *Drosophila* neurogenic locus mastermind encodes a nuclear protein unusually rich in amino acid homopolymers.** *Genes Dev* 1990, **4**:1688-1700.
27. Artavanis-Tsakonas S, Rand MD, Lake RJ: **Notch signaling: Cell fate control and signal integration in development.** *Science* 1999, **284**:770-776.
28. Helms W, Lee H, Ammerman M, Parks AL, Muskavitch MA, Yedvobnick B: **Engineered truncations in the *Drosophila* mastermind protein disrupt Notch pathway function.** *Dev Biol* 1999, **215**:358-374.
29. Petcherski AG, Kimble J: **Mastermind is a putative activator for Notch.** *Curr Biol* 2000, **10**:471-473.
30. Fryer CF, Lamar E, Turbachova I, Kintner C, Jones KA: **Mastermind mediates chromatin-specific transcription and turnover of the Notch enhancer complex.** *Genes Dev* 2002, **16**:1397-1411.
31. Lai EC: **Notch signalling: control of cell communication and cell fate.** *Development* 2004, **131**:965-973.
32. Bray SJ: **Notch signalling: a simple pathway becomes complex.** *Nat Rev Mol Cell Biol* 2006, **7**:678-689.
33. Oyama T, Harigaya K, Muradil A, Hozumi K, Habu S, Oguro H, Iwama A, Matsuno K, Sakamoto R, Sato M, Yoshida N, Kitagawa M: **Mastermind-1 is required for Notch signal-dependent steps in lymphocyte development in vivo.** *Proc Natl Acad Sci USA* 2007, **104**:9764-9769.
34. Chan SM, Weng AP, Tibshirani R, Aster JC, Utz PJ: **Notch signals positively regulate activity of the mTOR pathway in T-cell acute lymphoblastic leukemia.** *Blood* 2007, **110**:278-286.
35. Garg V, Muth AN, Ransom JF, Schluterman MK, Barnes R, King IN, Grossfeld PD, Srivastava D: **Mutations in NOTCH1 cause aortic valve disease.** *Nature* 2005, **437**:270-274.
36. Hamaratoglu F, Willecke M, Kango-Singh M, Nolo R, Hyun E, Tao C, Jafar-Nejad H, Halder G: **The tumour-suppressor genes *NF2/Merlin* and *Expanded* act through Hippo signalling to regulate cell proliferation and apoptosis.** *Nat Cell Biol* 2006, **8**:27-36.
37. Okada T, You L, Giancotti F: **Shedding light on Merlin's wizardry.** *Trends Cell Biol* 2007, **17**:222-229.
38. McClatchey AI, Giovannini M: **Membrane organization and tumorigenesis: the *NF2* tumor suppressor, Merlin.** *Genes Dev* 2005, **19**:2265-2277.
39. Jünger MA, Rintelen F, Stocker H, Wasserman JD, Végh M, Radimerski T, Greenberg ME, Hafen E: **The *Drosophila* forkhead transcription factor *FOXO* mediates the reduction in cell number associated with reduced insulin signaling.** *J Biol* 2003, **2**:20.
40. Puig O, Marr MT, Ruhf ML, Tjian R: **Control of cell number by *Drosophila* *FOXO*: downstream and feedback regulation of the insulin receptor pathway.** *Genes Dev* 2003, **17**:2006-2020.
41. Shingleton AW, Das J, Vinicius L, Stern DL: **The temporal requirements for insulin signaling during development in *Drosophila*.** *PLoS Biol* 2005, **3**:e289.
42. Nijhout HF: **The control of growth.** *Development* 2003, **130**:5863-5867.
43. Caldwell PE, Walkiewicz M, Stern M: **Ras activity in the *Drosophila* prothoracic gland regulates body size and developmental rate via ecdysone release.** *Curr Biol* 2005, **15**:1785-1795.
44. Mirth C, Truman JW, Riddiford LM: **The role of the prothoracic gland in determining critical weight for metamorphosis in *Drosophila melanogaster*.** *Curr Biol* 2005, **15**:1796-1807.
45. Colombani J, Bianchini L, Layalle S, Pondeville E, Dauphin-Villeman C, Antoniewski C, Carré C, Noselli S, Léopold P: **Antagonistic actions of ecdysone and insulins determine final size in *Drosophila*.** *Science* 2005, **310**:667-670.
46. Giot L, Bader JS, Brouwer C, Chaudhuri A, Kuang B, Li Y, Hao YL, Ooi CE, Godwin B, Vitols E, Vijayadamar G, Pochart P, Machineni H, Welsh M, Kong Y, Zerhusen B, Malcolm R, Varrone Z, Collis J, Minto M, Burgess S, McDaniel L, Stimpson E, Spriggs F, Williams J, Neurath K, Ioine N, Agee M, Voss E, Furtak K, Renzulli R, Aanensen N, Carrolla S, Bickelhaupt E, Lazovatsky Y, DaSilva A, Zhong J, Stanvyn CA, Finley RL Jr, White KP, Braverman M, Jarvie T, Gold S, Leach M, Knight J, Shimkets RA, McKenna MP, Chant J, Rothberg JM: **A protein interaction map of *Drosophila melanogaster*.** *Science* 2003, **302**:1727-1736.
47. James AC, Partridge L: **Thermal evolution of the rate of larval development in *Drosophila melanogaster* in laboratory and field populations.** *J Evol Biol* 1995, **8**:315-330.
48. Folguera G, Ceballos S, Spezzi L, Fanara JJ, Hasson E: **Clinal variation in developmental time and viability, and the response to thermal treatments in two species of *Drosophila*.** *Biol J Linn Soc* in press.
49. Stillwell RC, Fox CW: **Complex patterns of phenotypic plasticity: interactive effects of temperature during rearing and oviposition.** *Ecology* 2005, **86**:924-934.

50. Bateman JM, McNeill H: **Temporal control of differentiation by the insulin receptor/Tor pathway in *Drosophila*.** *Cell* 2004, **119**:87-96.
51. Li T, White KP: **Tissue-specific gene expression and ecdysone-regulated genomic networks in *Drosophila*.** *Developmental Cell* 2003, **5**:59-72.
52. Kyriacou CP, Oldroyd M, Wood J, Sharp M, Hill M: **Clock mutations alter developmental timing in *Drosophila*.** *Heredity* 1990, **64**:395-401.
53. Moss EG: **Heterochronic genes and the nature of developmental time.** *Current Biology* 2007, **17**:425-434.
54. Norga KK, Gurganus MC, Dilda CL, Yamamoto A, Lyman RF, Patel PH, Rubin GM, Hoskins RA, Mackay TF, Bellen HJ: **Quantitative analysis of bristle number in *Drosophila* mutants identifies genes involved in neural development.** *Curr Biol* 2003, **13**:1388-1397.
55. Harbison ST, Yamamoto AH, Fanara JJ, Norga KK, Mackay TF: **Quantitative trait loci affecting starvation resistance in *Drosophila melanogaster*.** *Genetics* 2004, **166**:1807-1823.
56. Edgar B: **How flies get their size: genetics meets physiology.** *Nat Rev Genet* 2006, **7**:907-916.
57. Bardin AJ, Le Borgne R, Schweisguth F: **Asymmetric localization and function of cell-fate determinants: a fly's view.** *Curr Opin Neurobiol* 2004, **14**:6-14.
58. Markow T, O'Grady P: ***Drosophila*: a guide to species identification and use.** Academic Press; 2005.
59. Paranjpe DA, Anitha D, Chandrashekar MK, Joshi A, Sharma VK: **Possible role of eclosion rhythm in mediating the effects of light-dark environments on pre-adult development in *Drosophila melanogaster*.** *BMC Developmental Biology* 2005, **5**:5.
60. Leips J, Mackay TF: **Quantitative trait loci for life span in *Drosophila melanogaster*: interactions with genetic background and larval density.** *Genetics* 2000, **155**:1773-1788.
61. Carlborg O, Jacobsson L, Ahgren P, Siegel P, Andersson L: **Epistasis and the release of genetic variation during long-term selection.** *Nat Genet* 2000, **38**:418-420.
62. Le Rouzic A, Carlborg O: **Evolutionary potential of hidden genetic variation.** *Trends Ecol Evol* 2008, **23**:33-37.
63. Lukacsovich T, Asztalos Z, Awano W, Baba K, Kondo S, Niwa S, Yamamoto D: **Dual-tagging gene trap of novel genes in *Drosophila melanogaster*.** *Genetics* 2001, **157**:727-742.
64. Robertson A: **The sampling variance of the genetic correlation coefficient.** *Biometrics* 1959, **15**:469-485.
65. Ungerer MC, Halldorsdottir SS, Purugganan MD, Mackay TF: **Genotype-environment interactions at quantitative trait loci affecting inflorescence development in *Arabidopsis thaliana*.** *Genetics* 2003, **165**:353-365.
66. The Gene Ontology Consortium: **Gene Ontology: tool for the unification of biology.** *Nature Genet* 2000, **25**:25-29.
67. Al-Shahrour F, Minguez P, Vaquerizas JM, Conde L, Dopazo J: **BABELOMICS: a suite of web tools for functional annotation and analysis of groups of genes in high-throughput experiments.** *Nucleic Acids Res* 2005, **33**:460-464.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

